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DOI:

[10.1002/cam4.871](https://doi.org/10.1002/cam4.871)

Document Version

Publisher's PDF, also known as Version of record

[Link to publication record in King's Research Portal](#)

Citation for published version (APA):

Shanmugalingam, T., Bosco, C. T., Ridley, A., & Van Hemelrijck, M. (2016). Is there a role for IGF-1 in the development of second primary cancers? *Cancer Medicine*. <https://doi.org/10.1002/cam4.871>

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REVIEW

Is there a role for IGF-1 in the development of second primary cancers?

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Keywords

Breast cancer, colorectal cancer, IGF-1, lung cancer, prostate cancer, second primary cancer

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Funding Information

This research was supported by the Experimental Cancer Medicine Centre at King's College London, Cancer Research UK (AJR), and also by the National Institute for Health Research (NIHR) Biomedical Research Centre based at Guy's and St Thomas' NHS Foundation Trust and King's College London.

Received: 9 October 2015; Revised: 8 June 2016; Accepted: 24 June 2016

doi: 10.1002/cam4.871

Abstract

Cancer survival rates are increasing, and as a result, more cancer survivors are exposed to the risk of developing a second primary cancer (SPC). It has been hypothesized that one of the underlying mechanisms for this risk could be mediated by variations in insulin-like growth factor-1 (IGF-1). This review summarizes the current epidemiological evidence to identify whether IGF-1 plays a role in the development of SPCs. IGF-1 is known to promote cancer development by inhibiting apoptosis and stimulating cell proliferation. Epidemiological studies have reported a positive association between circulating IGF-1 levels and various primary cancers, such as breast, colorectal, and prostate cancer. The role of IGF-1 in increasing SPC risk has been explored less. Nonetheless, several experimental studies have observed a deregulation of the IGF-1 pathway, which may explain the association between IGF-1 and SPCs. Thus, measuring serum IGF-1 may serve as a useful marker in assessing the risk of SPCs, and therefore, more translational experimental and epidemiological studies are needed to further disentangle the role of IGF-1 in the development of specific SPCs.

Introduction

Within the last 40 years, cancer survival rates have doubled in the UK [1], mainly due to advances in treatment, and the increased detection of cancer at an early stage [2]. In England and Wales, approximately 50% of adult cancer patients diagnosed in 2010 to 2011 are predicted to survive 10 years or more [1]. In the US, the 5-year relative survival rate for all cancers diagnosed in 2004–2010 was 68%, an increase from 49% in 1975–1977 [3]. Side-effects of cancer treatment and possible underlying etiological mechanisms, such as IGF-1 metabolism, are thought to be implicated in the development of second primary

cancers (SPCs). Therefore, identifying which cancer survivors have a high risk of developing SPCs is crucial.

It is well known that smoking [4, 5], obesity [6, 7], and insulin resistance [8, 9] are risk factors for the development of first primary cancers. However, the role of these risk factors in the development of SPCs in cancer survivors is less clear. There is some evidence that SPCs may be the result of genetic and hormonal risk factors [10–12], of late effects of chemo- and radiotherapy [2, 13], smoking and alcohol effects [14, 15], as well as nonmodifiable variables such as age and gender. For instance, a study based on the Swedish Family-Cancer Database concluded that, compared with the

general population, males and females diagnosed with an initial primary cancer were 1.3–1.6 times more likely to develop a second cancer, respectively [16].

Epidemiological evidence suggests that survivors of certain types of cancers have a higher risk of developing SPCs. For example, contralateral breast cancer is the most common SPC that develops in patients diagnosed with a first breast cancer, accounting for approximately 50% of all SPCs [17]. Furthermore, breast cancer has emerged as the most common solid cancer among female survivors of Hodgkin's lymphoma (diagnosed in childhood), which is largely due to the high-dose chest irradiation for Hodgkin's lymphoma [2, 18]. In addition, urological cancers (bladder, kidney, testes, and penile cancers) are consistently more prevalent among men with prostate cancer [19]. Indeed, it has been hypothesized that variations in the insulin-like growth factor (IGF) pathway, specifically IGF-1 and its binding protein 3 (IGFBP-3), could account for the increased risk of SPCs [20].

Recently, several studies have identified IGF-1 to be associated with an increased risk of developing a number of common cancers, including lung [21], breast [22], colorectal [23], and prostate [24]. Circulating levels of IGF-1 have been linked to the development of SPCs in men with head and neck squamous cell carcinoma [20]. However, so far a role of IGF-1 in development of SPCs following diagnosis of prostate cancer, breast cancer, colorectal cancer, or lung cancer has not been analyzed. In addition, a disorder known as Laron syndrome which is associated with low circulating levels of IGF-1 and IGFBP-3 [25] are protected from developing cancer, but instead can develop diabetes and cardiovascular disease [26].

With the increase in number of cancer survivors, the long-term health outcomes of this population need to be carefully examined. Approximately, one in five cancers is diagnosed in those with a previous diagnosis of cancer, and hence, these “second primary cancers” are a leading cause of morbidity and mortality among cancer survivors [27]. It is therefore of interest to investigate the role of IGF-1 in the development of various SPCs as it can help us understand the potential underlying mechanism for carcinogenesis. This review therefore aims to identify whether IGF-1 plays a role in the development of SPCs, by assessing epidemiological evidence available to date.

Literature Review

We used a computerized literature search database (PubMed and EMBASE) to identify full text and abstract studies of English language, using human subjects and published between the years 1999 and 2015. Searches were performed with and without the Medical Subject Heading (MeSH) terms for “cancer”, “breast cancer”, “lung cancer”,

“prostate cancer”, “colorectal cancer”, and “meta analysis”, combined with the keywords “second primary cancer” and “IGF 1”. All references of the selected articles were checked using hand searches.

IGF-1 in First Primary Cancers

This section provides an overview of evidence for the emerging role of IGF-1 in the development of first primary cancers, with a focus on epidemiological studies (Table 1) as well as experimental studies investigating the underlying biological mechanisms.

IGF-1 is a single-chain polypeptide growth factor [28–30] that is related to insulin and IGF-2 [31]. IGF-1 stimulates cell growth, proliferation, and differentiation, and is essential for normal organismal growth and development [32, 33]. IGF-1 binds to the insulin-like growth factor 1 receptor (IGF-1R), which is a tyrosine kinase receptor [34]. IGF-1 has a higher binding affinity than IGF-2 for IGF-1R. IGF-1R initiates a cascade of downstream signal transduction pathways known to be involved in cell growth, proliferation, and cancer, including Ras/Raf/ERK and PI3K/Akt/mTOR [35]. The majority of IGF-1 found in the circulation is produced by the liver, functioning as an endocrine hormone. IGF-1 is also produced in other organs where autocrine or paracrine mechanisms have a role [36]. Ample evidence indicates that IGF-1 and IGF-1R are important for growth and survival of cancer cells [37, 38] (Fig. 1). The expression of the IGF-1 gene is primarily regulated by growth hormone (GH), and to a smaller extent by various other hormones [35]. By contrast, IGF-1 that is synthesized locally in an autocrine or paracrine manner may stimulate growth of some cancers [36]. The circulating levels of IGF-1 change markedly with age, peaking at puberty, and slowly declining with increasing age; this fluctuation is regulated by GH, which itself has mitogenic and proliferative properties [35]. However, in other cell types, for example, cartilage cells, the growth-stimulating effects of IGF-1 are GH-independent [39]. Furthermore, GH deficiency is the most common disorder seen in survivors of childhood cancer, and there are concerns regarding its use in treating cancer survivors as it might increase the risk of SPCs [40]. Although IGF-1 possesses antiapoptotic, cell survival, and transforming activities, it is not classed as an oncogene.

Breast cancer

Findings to date on the role of IGF-1 in breast cancer development vary depending on the study. An early case-control study conducted in 1993 demonstrated that circulating levels of IGF-1 were higher in women with breast cancer compared to women without breast cancer [41].

Table 1. Studies of cancer risk related to IGF-1 level.

Author (Year)	Control (n)	Cases (n)	Cancer risk related to IGF-1 level	Reference
Breast cancer				
Peyrat (1993)	92	44	Median concentrations: 26 ng/mL (BCa) versus 20 ng/mL (controls)	[38]
Endogenous Hormones and Breast Cancer Collaborative Group (2010)	1839	1032	OR for BCa in the highest versus lowest fifth of IGF1 concentration was 1.28 (95% CI: 1.14–1.44; $P < 0.0001$)	[39]
Rinaldi (2006)	312	202	Highest versus lowest quintile OR 1.38 (95% CI: 1.02–1.86; $P = 0.01$) for women who develop breast cancer after 50 years of age	[40]
Kaaks (2014)	259	193	OR=1.41 (95% CI: 1.01–1.98; $P = 0.01$ for the highest versus lowest quartile, for ER+ breast tumors overall (pre- and postmenopausal women combined)	[41]
Baglietto (2007)	4296 ¹	119	HR for BCa comparing the fourth with the first quartiles was 1.20 (95% CI: 0.87–1.65).	[42]
	1954 ² versus 736	68 versus 9	HR for BCa in older women comparing the fourth with the first quartiles (+60 years) was 1.61 (95% CI: 1.04–2.51) versus 0.60 (95% CI: 0.25–1.45) for younger women (<50 years)	
Renehan (2004)	Meta-analysis of 4 studies	Meta-analysis of 4 studies	High concentrations of IGF-1 were associated with an increased risk of premenopausal BCa (OR comparing 75th with 25th percentile 1.65, 95% CI: 1.26–2.08; $P < 0.001$)	[44]
Shi (2004)	1306	779	Premenopausal women: Nearly 40% increase in BCa risk among those who had higher IGF-1 in the circulation (overall OR 1.39, 95% CI: 1.16–1.66).	[45]
	1552	911	No association in postmenopausal women (overall OR 0.93, 95% CI: 0.80–1.10).	
Sugumar (2004)	1471	764	Subjects with higher circulating levels of IGF-1 had increased risk of premenopausal BCa with an OR of 1.74 (95% CI: 0.97–3.13; $P = 0.06$)	[46]
Schernhammer (2006)	158	79	RR for top versus bottom quartile of IGF-1 was 0.98 (95% CI: 0.69–1.39; $P = 0.77$)	[47]
Hankinson (1998)	92	46	Postmenopausal women: No association between IGF-1 concentrations and BCa risk (top vs. bottom quintile of IGF-1, RR = 0.85 [95% CI: 0.53–1.39]).	[22]
	35	35	RR of BCa among premenopausal women by IGF-1 concentration (top vs. bottom tertile) was 2.33 (95% CI: 1.06–5.16; $P = 0.08$)	
Lung cancer				
Ahn (2006)	101	38	OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 0.69 (95% CI: 0.41–1.15); $P = 0.26$	[51]
London (2002)	159	51	OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 0.73 (95% CI: 0.43–1.24); $P = 0.80$	[53]
Lukanova (2001)	47	23	OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 0.79 (95% CI: 0.29–2.19); $P = 0.53$	[54]

(Continued)

Table 1. Studies of cancer risk related to IGF-1 level. (Continued)

Author (Year)	Control (n)	Cases (n)	Cancer risk related to IGF-1 level	Reference
Morris (2006)	11,072	843	Meta-analysis: OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 1.02 (95% CI: 0.80–1.31); $P = 0.64$	[55]
Yu (1999)	54	74	High plasma levels of IGF-1 were associated with an increased risk of LCa (OR = 2.06; 95% CI: 1.19–3.56; $P = 0.01$)	[21]
Chen (2009)	Meta-analysis of 6 studies	Meta-analysis of 6 studies	Pooled OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 0.87 (95% CI: 0.60–1.13)	[56]
Cao (2012)	Meta-analysis of 6 studies	Meta-analysis of 6 studies	OR for LCa risk by IGF-1 concentrations (highest vs. lowest quartile) was 1.05 (95% CI: 0.80–1.37); $P = 0.74$	[57]
Prostate cancer Mantzoros (1997)	52	51	Increment of 60 ng mL corresponded to an OR of 1.91 (95% CI: 1.00–3.73; $P = 0.05$)	[62]
Colorectal cancer Nomura (2003)	282	177 (colon cancer) 105 (rectal cancer)	Weakly positive association of IGF-I with colon cancer. Colon cancer cases in third (IGF-1 of 137–174 ng/mL) and fourth quartile (IGF-1 > 174 ng/mL) had increased risk compared with controls (OR of 2.2 and 1.8, respectively)	[68]
Palmqvist (2002)	336	110 (colon cancer) 58 (rectal cancer)	No association of IGFI with rectal cancer Increase in colon cancer risk with increasing levels of IGF-1 (OR of 2.30 and 2.66 for third and fourth quartile, respectively) Rectal cancer risk was inversely related to levels of IGF-1 (OR of 0.33 and 0.33 for third and fourth quartile, respectively)	[69]
Tripkovic (2007)	52	52	Increase in IGF-1 level was followed by a 3.15-fold increased risk for developing colon cancer with levels of IGF-1 > 310 ng/mL, whereas twice as many controls exhibited levels of IGF-1 < 107 ng/mL	[70]
Ma (1999)	318	193	Men in the highest quintile for IGF-I had an increased risk of colorectal cancer compared with men in the lowest quintile (RR = 2.51; 95% CI: 1.15–5.46; $P = 0.02$)	[71]
Kaaks (2000)	200	102	Colorectal cancer risk showed a modest but statistically nonsignificant positive association with levels of IGF-I	[72]

BCa, breast cancer; OR, odds ratio; CI, confidence intervals; HR, hazard ratios; RR, relative risk; LCa, lung cancer.

¹Breast cancer cases and person-years calculated from the 2284 women with IGF-I measured.

²Breast cancer cases and person-years.

Since then, several epidemiological studies have reported that higher circulating levels of IGF-1 are associated with an increased risk of breast cancer [22, 42–45]. This may arise as higher levels of IGF-1 are associated with acceleration of early carcinogenesis [36]. More recently, three

meta-analyses demonstrated positive associations between IGF-1 and risk of breast cancer among premenopausal, but not postmenopausal women [46–48]. A study by Schernhammer et al. aims to explain these observations by showing that premenopausal women with high IGF-1

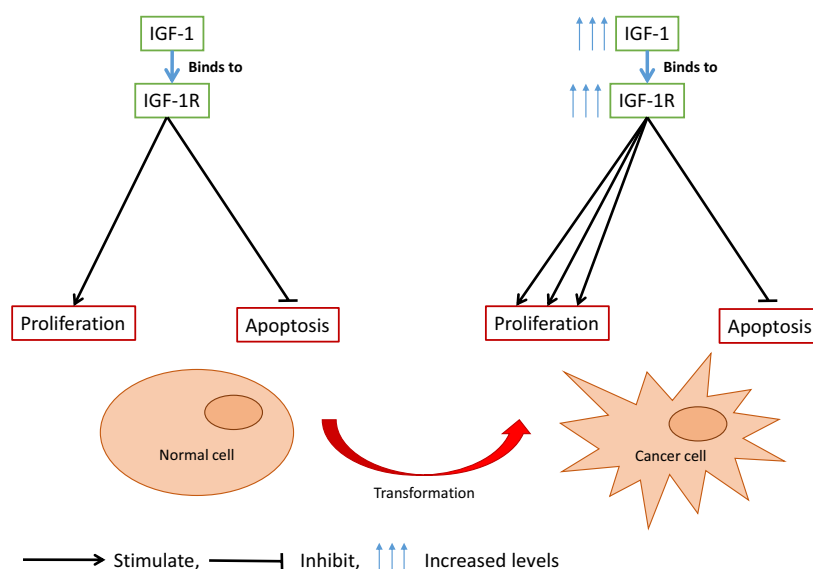


Figure 1. The effects of IGF-1 and IGF-1R on normal and cancerous cells.

levels were at risk of higher IGF-1R activation in mammary epithelial cells, which is suggested to increase survival of these cells with accumulating DNA damage, thereby facilitating stepwise carcinogenesis [49]. These results may indicate the importance of IGF-1 levels in younger women in early life or its possible interaction with other hormones such as estradiol [36] and growth hormone [39]. In contrast to these studies, a large prospective study pooling two Swedish cohorts found no association between circulating IGF-1 and risk of breast cancer, regardless of menopausal status [50]. It is unclear why there is such a discrepancy in study findings. However, differences in findings may be due to timing of blood sampling, the patient cohort or the subset of breast cancer. Prospective studies are advantageous over retrospective studies since blood samples to measure IGF-1 levels are collected before the clinical diagnosis of cancer and hence reverse causation (i.e., effects of an undiagnosed cancer on levels of IGF-1) is less likely to play a role [50].

Estrogen plays an important role in the etiology of breast cancer, and there are experimental studies reporting cross-talk between IGF-1 and the estrogen receptor (ER) in mammary cells, possibly through synergistic effects that contribute to breast carcinogenesis [44, 51]. Stewart et al. showed that estrogen increases IGF-1 binding and IGF-1R mRNA levels in the estrogen-sensitive MCF-7 cell line by 7- and 6.5-fold, respectively [52]. This suggests that one potential mechanism by which estrogen stimulates breast cancer cell proliferation may involve sensitization of IGF-1 [52].

Thus far, epidemiological evidence overall suggests a positive association between IGF-1 and breast cancer risk,

particularly in premenopausal women. Moreover, experimental evidence suggests that a link between IGF-1 and estrogen may explain this positive association, but perhaps only in breast cancers that express the estrogen receptor.

Lung cancer

Studies to date have investigated the association between IGF-1 and lung cancer. Several studies have shown that circulating IGF-1 levels were not associated with an increased risk of lung cancer [53–57]. One case–control study found a positive association between IGF-1 and risk of lung cancer (OR: 2.06; 95% CI: 1.19–3.56) [21]. Furthermore, this study identified that the levels of IGF-1 and IGF-2 in plasma were not influenced by cigarette smoking [21].

IGFBP-3 is the main IGF-1-binding protein in blood. IGFBP-3 is generally considered to act as a tumor suppressor gene by reducing the ability of IGF-1 to promote cell survival and proliferation [58]. Although epidemiological studies overall found no association for IGF-1, a reduced risk of lung cancer is reported with higher circulating levels of IGFBP-3, when comparing the highest quartile versus lowest quartile of IGFBP-3 in a Chinese prospective study (OR: 0.50, 95% CI: 0.25–1.02) [55]. Moreover, several meta-analyses have also reported an inverse association between IGFBP-3 and risk of lung cancer [58, 59].

It is possible that both IGF-1 and IGFBP-3 contribute to the development of lung cancer. Cell culture studies have found that lung cancer cell lines, regardless of their

histological subtypes, have the capacity to express IGF-1 and its binding protein, IGFBP-3, both in tumors and blood [60, 61].

Thus, until now, there is little evidence for a link between IGF-1 and lung cancer risk, but an inverse association between IGFBP-3 and lung cancer risk has been observed. These epidemiological observations are consistent with experimental data, which demonstrates that IGFBP-3 block the mitogenic and antiapoptotic effects of IGF-1 on lung cancer cells [21, 55].

Prostate cancer

Associations between prostate cancer and IGF-1 have been studied extensively, and consistently show a positive association. Since 1993, it has been investigated whether higher circulating IGF-1 levels are associated with an increased risk of prostate cancer [62, 63]. Early studies failed to demonstrate an association between IGF-1 and prostate cancer risk. The first significant positive association between IGF-1 and prostate cancer was examined in a case-control study by Mantzoros et al. By comparing men with prostate cancer to healthy controls, the odds ratio per 60 ng/mL increment in circulating levels of IGF-1 was 1.91 (95% CI: 1.00–3.73) [64]. Furthermore, the authors also mentioned that this association is further reinforced by the lack of association between IGF-1 and benign prostatic hyperplasia.

IGF-1 is known to stimulate the growth of prostate cancer cells by inducing cell proliferation and inhibiting apoptosis [65]. The effect of IGF-1 on prostate cancer cell lines has been extensively explored. For example, in

vivo studies have shown significantly reduced proliferation rates in PC-3 prostate cancer cell lines in IGF-1-deficient hosts, compared to control hosts [66]. Exogenous IGF-1 increased the invasive potential of the DU145 prostate cancer cell line, which was dependent on IGF-1R, the ERK MAPK pathway, and the PI3K pathway [67]. Furthermore, prostate cancer epithelial cells can stimulate their own growth by synthesizing and responding to IGF-1 in an autocrine manner (Fig. 2), as opposed to paracrine signaling [68].

Overall, epidemiological and experimental evidence to date suggest a positive association between circulating IGF-1 and prostate cancer risk.

Colorectal cancer

Overall, studies have provided data showing that colorectal cancer is positively associated with IGF-1 levels [23, 69–72]. Early studies in the late 1990s, suggested that high circulating IGF-1 concentrations are associated with an increasing risk of colorectal cancer [23, 73]. Nomura et al. evaluated the association between IGF-1 and colon and rectal cancer separately [69]. They showed a higher risk of colon cancer for those with IGF-1 levels in the third (137–174 ng/mL) and fourth quartiles (IGF-1 > 174 ng/mL) as compared to the controls (OR of 2.2 and 1.8, respectively). There was no association between IGF-1 and rectal cancer. However, another study found a decreased risk of rectal cancer with high levels of circulating IGF-1 [70]. The authors commented that this may be due to rectal cancer presenting at an earlier stage than colon cancer, which may have masked the association with IGF-1

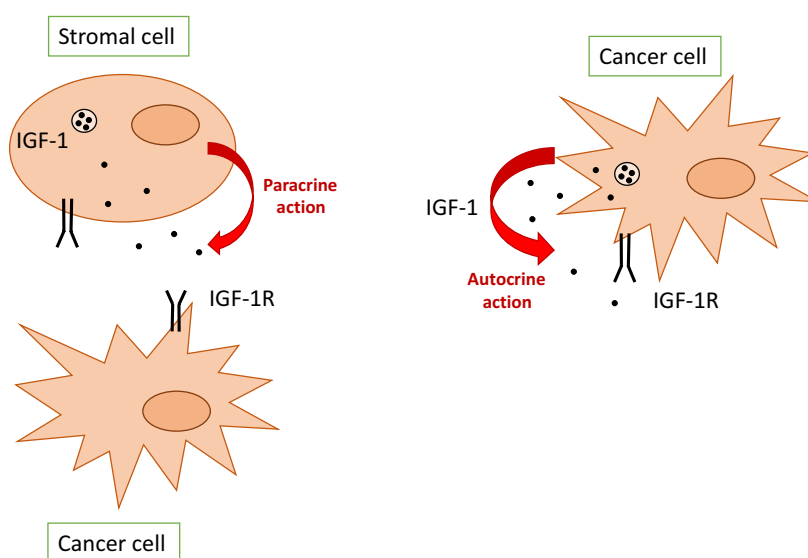


Figure 2. The autocrine and paracrine actions of IGF-1.

levels [70]. On the other hand, data from two nested case-control study showed no statistically significant association between IGF-1 and colorectal cancer risk [74, 75].

The epidemiological data on IGF-1 and colorectal cancer is supported by laboratory evidence. The IGF-1R is expressed in both normal colonic mucosa and human colorectal cancers [70]. IGF-1 has been speculated to cause proliferation of colorectal cancer cells and promote over-expression of the IGF-1R in several studies, with no uniform consensus [76–79]. Furthermore, in acromegaly, a condition that is associated with high IGF-1 levels, studies have shown that there is increased proliferation of normal colonic epithelium with an increased risk of developing colorectal adenomas and cancers in acromegaly patients [80, 81].

In colorectal cancer, the circulating levels of IGF-1 is particularly influenced by nutritional status. Therefore, further research is required to investigate the role of obesity, ethnicity, and dietary habits possibly as confounders to IGF-1 and colorectal cancer risk.

IGF-1 in Second Primary Cancers

This section summarizes studies on the emerging role of IGF-1 in the development of SPCs with a focus on the definition of SPC as well as their potential link with IGF-1 in the case of patients with primary breast, lung, prostate, and colorectal cancer.

SPCs are defined as malignant tumors diagnosed at the same time as the primary tumor or later, which are in a different organ [82] and are not a metastasis or recurrence of the original primary cancer [83].

Breast cancer

The risk of developing a second primary breast cancer in a patient diagnosed with a first breast cancer has been studied extensively. It has been shown that in women with breast cancer, the risk of developing a new primary breast cancer in the contralateral breast is much higher than for healthy women developing a first breast cancer [84, 85]. Only a small portion of this large risk is attributable to effects of treatment: lifestyle and genetic factors also need to be taken into account [84], and possibly the role of IGF-1 in increasing breast cancer risk.

Breast cancer is also a common SPC in itself. For instance, it is the most common SPC that develops in young women treated for Hodgkin's lymphoma with supradiaphragmatic irradiation [86]. The estimates of cumulative risks of developing breast cancer ranged from 35% by 40 years of age [87] to 48% by 40 years of age [88] after treatment. Much of this variation may be due to the differences in the age of diagnosis of Hodgkin's

lymphoma during radiation treatment, or the duration and dose of treatment [88, 89]. Even in the absence of treatment effects, in general, younger women are at a greater risk of developing a second primary breast cancer than older women [90, 91]. For instance, a cohort study by Hancock et al. which reviewed women treated for Hodgkin's disease between 1961 and 1990 (mean follow-up, 10 years) concluded that the greatest risk was seen in young women treated before the age of 15 years (RR: 136, 95% CI: 34–371), with a significant decline in relative risk with advancing age (above 30 years of age, RR: 0.7, 95% CI: 0.2–1.8) [92].

From a biological point of view, the pubertal growth of the mammary gland is mediated predominantly by the actions of IGF-1 and GH via estrogen [93]. It is postulated that this mechanism may make younger women more prone to developing a secondary breast cancer due to the increased levels of both IGF-1 and estrogen during puberty, and the promoting effects of IGF-1 [41, 94]. According to the evidence from *in situ* hybridization, in breast cancer, IGF-1 is predominantly expressed in the stromal cells (mainly fibroblasts) and very rarely in the breast epithelium [28, 95]. This supports the concept of a paracrine role of IGF-1 in breast cancer (Fig. 2). It is possible that there is also an endocrine role, given that circulating IGF-1 in the bloodstream is implicated in the malignant transformation of breast tissue [28, 43, 96]. IGF-1 functions to protect breast cancer cells from apoptosis and induces survival [28], suggesting that locally synthesized IGF-1 influences the growth of human breast cancer cells. It can therefore be hypothesized that IGF-1 produced by the stromal cells is increased in breast cancer (Fig. 2). This may then promote growth of a second primary breast cancer by entering the blood stream and acting in an endocrine fashion [94, 97].

Lung cancer

The lungs are often regarded as one of the most common organs to develop a SPC [11, 98, 99]. Common causes of a secondary lung cancer include a resected primary lung cancer, treatment-related complications in breast cancer and head and neck cancers [99–101], as well as continued smoking [102, 103]. In a US study, Johnson reported a 2–14% risk of developing a second lung cancer per person per year, with the risk increasing from twofold to sevenfold after 10 years of initial lung cancer diagnosis [104].

Head and neck cancer patients are at an increased risk of developing lung cancer with a standardized incidence ratio (SIR) of 3.75 (95% CI: 3.01–4.62) [98]. In addition, according to data from the Surveillance, Epidemiology, and End Results (SEER) registries, approximately 5% of

breast cancer survivors are diagnosed with a second primary lung cancer [11]. Additionally, treatment of breast cancer patients with radiation postmastectomy has been shown to approximately double the risk of second primary lung cancer, especially in the ipsilateral lung [99]. The rates of second primary lung cancer among women diagnosed with breast cancer before the age of 50 years is rising significantly, with the increase being as early as 1 year after breast cancer diagnosis [105]. Radiotherapy treatment for breast cancer seems unlikely as the sole cause of this rise (see methodological section) because a long-term latency period (5–10 years) is usually associated with radiotherapy treatment [105]; proteins or hormones such as IGF-1 should also be considered as predisposing factors.

When considering the biological effects of IGF-1 in second primary lung cancer, it has been shown that lung mesenchymal cells locally synthesize IGF-1 which acts on the bronchial epithelium in a paracrine manner [56]. A possible mechanism explaining the role of IGF-1 in second primary lung cancer is that higher IGF-1 levels detected in lung cancer are probably regulated by the levels of tissue-derived IGF-1, but not circulating IGF-1 [106].

Prostate cancer

The risk of all SPCs following a diagnosis of a first primary prostate cancer has been studied with variable results. In 1999, a Swiss study based on data collected from the Cancer Registries of the Swiss Cantons of Vaud and Neuchâtel investigated the risk of SPCs in prostate cancer survivors between 1974 and 1994. They found a significantly reduced incidence rate of all cancers in men diagnosed with prostate cancer, compared with the general population (SIR: 0.7, 95% CI: 0.6–0.8) [107]. More recently, in 2014, a cohort study from the Swiss Canton of Zurich investigated the risk of SPCs in prostate cancer survivors between 1980 and 2010 [19]. They found an increased risk of SPCs among men with prostate cancer, compared to the general population (SIR: 1.11, 95% CI: 1.06–1.17). The inconsistency between these two studies may be due to the diagnosis of prostate cancer at an advanced stage with shorter survival in the earlier years of study [19]. Therefore, the chance of developing a SPC was lower than what it is currently, when prostate cancer is generally diagnosed at a less advanced stage.

When looking at specific cancer types, Davies et al. reported that survivors of prostate cancer had a 40% lower risk of developing a SPC compared to the general male US population; the risk was lower for leukemias and cancers of the oral cavity, stomach, colon, liver, lung, and pancreas [108]. However, they observed a higher risk of developing bladder [109, 110], renal, and endocrine

cancers [111, 112]; this seems to be influenced by pelvic radiation therapy for prostate cancer [108]. Moreover, diagnostic bias is thought to play a role due to anatomy. However, Chrouser et al. did not observe an increased risk of bladder cancer after radiotherapy for prostate cancer [113], and there are some uncertainties in relation to the possible mechanism for the lack of association observed in this study. It is possible that there may have been an increased risk in this study that was not detected due to a short mean follow-up period of 7.1 years or likelihood of underreporting SPCs. Based on the current evidence, it seems that the risk of developing a SPC after prostate cancer is higher, particularly for other urological cancers.

Prostate cancer is also commonly observed as a SPC in itself. Kok et al. concluded that in the first year following a first cancer diagnosis, male cancer survivors have a 30% increased risk of developing prostate cancer as a SPC, partly due to increased diagnostic activity of the urological organs or incidental finding following health check-ups [114]. Other studies have also shown an excess risk of developing prostate cancer as a SPC after a diagnosis of a bladder cancer as a first cancer [115, 116]. In addition, cancer survivors diagnosed with a first primary urological cancer may request for prostate-specific antigen (PSA) testing as a consequence of anxiety or persisting urological symptoms [114]. Furthermore, survivors of melanoma are also at increased risk of developing prostate cancer [117].

In contrast to the biological mechanisms of breast and lung cancer, prostate cancer epithelial cells can stimulate their own growth by synthesizing and responding to IGF-1 [65, 68]. Furthermore, there is evidence that IGF-1 enhances the adhesion of prostate cancer cells and this promotes prostate cancer metastasis, possibly through the actions of IL-17 [118]. The potential data does not suggest a direct causative role for IGF-1 signaling in the progression and invasiveness of prostate cancer. The IGF-1 pathway activates a number of downstream signaling pathways, including the phosphatidylinositol-3 kinase (PI3-K) pathway, the protein kinase C pathway, the CREB pathway, and the mitogen-activated protein kinase (MAPK) pathway. These pathways contribute to prostate cancer through deregulation and constitutive activation of the pathway [67]. While the etiology of IGF-1 in second primary prostate cancer is unknown, it is plausible that those who develop prostate cancer may possess a common genetic, hormonal, or environmental factor that protects them from developing a SPC [119]. Prostate cancer survivors have a lower risk of developing cancers of the stomach, lung, and pancreas [108, 120], raising the question of whether these patients are “protected” against these malignancies, or whether it is simply that they are above the

age at which the risk of these tumors typical peak, which is at an earlier age [119].

Colorectal cancer

Several studies have demonstrated an increased risk of developing secondary colorectal cancer following radiotherapy exposure, in particular, rectal cancer following radiation for prostate cancer and colorectal cancer following abdominopelvic radiation for cervical cancer. Brenner et al. investigated the risk in prostate cancer patients who underwent radiotherapy or surgery and reported a significantly increased risk of rectal cancer in the radiotherapy group, particularly for long-term survivors, when comparing with the surgery group [121]. Furthermore, Baxter et al. observed a significant increase in the development of rectal cancer postradiation for prostate cancer [122]. However, radiation did not promote development of cancer in the remainder of the colon, suggesting that the effect of radiation is limited directly to irradiated tissue. In addition to prostate cancer patients, cervical cancer patients also seem to be at risk of developing colon cancer, as observed by Chatruvedi et al. [123].

In normal colonic tissue, IGF-1 binds with high affinity to the IGF-1R and activates specific insulin receptor substrates, which can modulate several downstream pathways involved in gene transcription, cell proliferation, and apoptosis [124]. Although the etiology of IGF-1 in second primary colorectal cancer is unknown, based on findings from normal colonic tissues, we can speculate about the potential complexity of this carcinogenic mechanism. With the exposure to radiotherapy, one hypothesis suggests that in individuals with higher IGF bioactivity, there is enhanced survival of partially transformed cells which leads to a larger pool of targets for subsequent “hits” initiating colorectal carcinogenesis via the process of stepwise carcinogenesis and malignant transformation. A second hypothesis suggests that the time needed for the progression of a fully transformed cell to fully developed cancer is inversely associated with IGF bioactivity [69].

Methodological considerations for epidemiological studies investigating the link between IGF-1 and risk of second primary cancers

In the clinical setting, it may be problematic to absolutely define whether the second tumor is in fact a SPC or a recurrence or a metastasis, and a definitive diagnosis may only be possible histologically. Whether the results showed in this review were strictly according to the standard are unclear, so therefore we need to consider the results with caution.

Even though several studies suggest a link between IGF-1 and development of SPCs, several methodological issues need to be considered when assessing these epidemiological findings. Firstly, diagnostic bias may occur when the SPC is the main outcome of interest, as it may be detected following a diagnostic intervention related to the first primary tumor [115]. Aside from diagnostic activity, treatment related to this first primary tumor may also increase the risk of developing a second primary tumor (e.g., chemotherapy and radiation therapy) [115, 125].

Secondly, when evaluating the effect of IGF-1 on SPCs, one has to consider sources of errors that cause misclassification of this biomarker. Nondifferential misclassification of IGF-1 may occur due to laboratory errors (e.g., specimen collection, processing, and storage) or changes in IGF-1 assays [126]. In addition, a single measurement of IGF-1 may not reflect the actual underlying levels. Repeated measurements would reflect long-term exposure and may be useful in the context of carcinogenesis [127]. Aside from misclassification of data for IGF-1, it is also possible to have misclassification related to the SPCs because it is not always possible from a pathological point of view to make a distinction between local recurrences, metastases, or a true SPC.

Thirdly, when studying the association between IGF-1 and SPCs, one has to be aware of potential confounders such as smoking or treatment. In the case of lung cancer, the effect of current and past smoking needs to be removed to maximize the efficiency of the study [56] as some studies have shown that smoking decreases the levels of IGF-1 [128], while others have found no relationship [129]. It is therefore possible that cigarette-related carcinogen exposure may overshadow the more subtle effects of IGF-1 on cancer development, which could explain the general lack of an association between IGF-1 and risk of lung cancer [54]. As a result, smoking may have an effect on IGF-1 levels as well as the risk of developing a SPC, but it is unlikely to be an intermediate in the pathway between IGF-1 and SPCs [130]. Furthermore, treatment received for the primary cancer may confound the association between IGF-1 and SPCs [114, 130]. Similarly to smoking, treatment may also alter the effect of IGF-1 as well as the risk of developing a SPC, but it is again unlikely to be an intermediate in the pathway between IGF-1 and SPCs [130].

Conclusion

In spite of a consistent positive observation between IGF-1 and risk of first primary cancers (especially breast, prostate, and colorectal), the evidence for the role of IGF-1 in the development of SPCs is less clear. Some of the evidence we gathered came from targeting the IGF system in cell

culture studies, and therefore, we need to see the results with caution on whether it can be compared to clinical situations. However, the relevant influences of these pathways in SPCs are unknown. This lack of an association may be partly explained by methodological issues. With respect to the biological pathway, there is consistent evidence for the mitogenic role of IGF-1 in carcinogenesis by increasing cell proliferation and inhibiting apoptosis. However, experimental studies highlight uncertainties regarding the role of IGF-1 in the development of SPCs. More observational studies are needed to further understand the role of IGF-1 in the development of specific SPCs, as well as to determine which pathways downstream of the IGF-1R are involved in this process.

Acknowledgments

This research was supported by the Experimental Cancer Medicine Centre at King's College London, Cancer Research UK (AJR) and also by the National Institute for Health Research (NIHR) Biomedical Research Centre based at Guy's and St Thomas' NHS Foundation Trust and King's College London. The views expressed are those of the author(s) and not necessarily those of the NHS, the NIHR, or the Department of Health.

Conflict of Interest

None declared.

References

- Morris, L. G., A. G. Sikora, S. G. Patel, R. B. Hayes, and I. Ganly. 2011. Second primary cancers after an index head and neck cancer: subsite-specific trends in the era of human papillomavirus-associated oropharyngeal cancer. *J. Clin. Oncol.* 29:739–746.
- Travis, L. B. 2006. The epidemiology of second primary cancers. *Cancer Epidemiol. Biomarkers Prev.* 15:2020–2026.
- Cancer Facts & Figures. 2015. Atlanta, Georgia: American Cancer Society; 2015; Available from: <http://www.cancer.org/acs/groups/content/@editorial/documents/document/acspc-044552.pdf> (accessed date: 7 February 2016).
- Sasco, A. J., M. B. Secretan, and K. Straif. 2004. Tobacco smoking and cancer: a brief review of recent epidemiological evidence. *Lung Cancer* 45(Suppl 2):S3–S9.
- Lee, P. N., B. A. Forey, and K. J. Coombs. 2012. Systematic review with meta-analysis of the epidemiological evidence in the 1900s relating smoking to lung cancer. *BMC Cancer* 12:385.
- Wolk, A., G. Gridley, M. Svensson, O. Nyren, J. K. McLaughlin, J. F. Fraumeni, et al. 2001. A prospective study of obesity and cancer risk (Sweden). *Cancer Causes Control* 12:13–21.
- Bianchini, F., R. Kaaks, and H. Vainio. 2002. Overweight, obesity, and cancer risk. *Lancet Oncol.* 3:565–574.
- Tsugane, S., and M. Inoue. 2010. Insulin resistance and cancer: epidemiological evidence. *Cancer Sci.* 101:1073–1079.
- Trevisan, M., J. Liu, P. Muti, G. Misciagna, A. Menotti, F. Fucci, et al. 2001. Markers of insulin resistance and colorectal cancer mortality. *Cancer Epidemiol. Biomark. Prev.* 10:937–941.
- Travis, L. B., C. S. Rabkin, L. M. Brown, J. M. Allan, B. P. Alter, C. B. Ambrosone, et al. 2006. Cancer survivorship–genetic susceptibility and second primary cancers: research strategies and recommendations. *J. Natl Cancer Inst.* 4:15–25.
- Mariotto, A. B., J. H. Rowland, L. A. Ries, S. Scoppa, and E. J. Feuer. 2007. Multiple cancer prevalence: a growing challenge in long-term survivorship. *Cancer Epidemiol. Biomark. Prev.* 16:566–571.
- Travis, L. B., W. Demark Wahnefried, J. M. Allan, M. E. Wood, and A. K. Ng. 2013. Aetiology, genetics and prevention of secondary neoplasms in adult cancer survivors. *Nature reviews. Clin. Oncol. (R. Coll. Radiol.)* 10:289–301.
- Rheingold, S. R., A. I. Neugut, and A. T. Meadows. 2003. Therapy-related secondary cancers. 6th ed. BC Decker. Medicine H-FC, editor.
- McLaughlin, V. H., A. Trentham-Dietz, J. M. Hampton, P. A. Newcomb, and B. L. Sprague. 2014. Lifestyle factors and the risk of a second breast cancer after ductal carcinoma in situ. *Cancer Epidemiol. Biomark. Prev.* 23:450–460.
- Li, X., and K. Hemminki. 2003. Familial and second lung cancers: a nation-wide epidemiologic study from Sweden. *Lung Cancer* 39:255–263.
- Dong, C., and K. Hemminki. 2001. Second primary neoplasms in 633,964 cancer patients in Sweden, 1958–1996. *Int. J. Cancer* 93:155–161.
- Gao, X., S. G. Fisher, and B. Emami. 2003. Risk of second primary cancer in the contralateral breast in women treated for early-stage breast cancer: a population-based study. *Int. J. Radiat. Oncol. Biol. Phys.* 15:1038–1045.
- Neglia, J. P., D. L. Friedman, Y. Yasui, A. C. Mertens, S. Hammond, M. Stovall, et al. 2001. Second malignant neoplasms in five-year survivors of childhood cancer: childhood cancer survivor study. *J. Natl Cancer Inst.* 93:618–629.
- Van Hemelrijck, M., A. Feller, H. Garmo, F. Valeri, D. Korol, S. Dehler, et al. 2014. Incidence of second malignancies for prostate cancer. *PLoS ONE* 9:e102596.

20. Wu, X., H. Zhao, K. A. Do, M. M. Johnson, Q. Dong, W. K. Hong, et al. 2004. Serum levels of insulin growth factor (IGF-I) and IGF-binding protein predict risk of second primary tumors in patients with head and neck cancer. *Clin. Cancer Res.* 10(12 Pt 1):3988–3995.
21. Yu, H., M. R. Spitz, J. Mistry, J. Gu, W. K. Hong, and X. Wu. 1999. Plasma levels of insulin-like growth factor-I and lung cancer risk: a case-control analysis. *J. Natl Cancer Inst.* 20:151–156.
22. Hankinson, S. E., W. C. Willett, G. A. Colditz, D. J. Hunter, D. S. Michaud, B. Deroo, et al. 1998 May. Circulating concentrations of insulin-like growth factor-I and risk of breast cancer. *Lancet* 9:1393–1396.
23. Ma, J., M. N. Pollak, E. Giovannucci, J. M. Chan, Y. Tao, C. H. Hennekens, et al. 1999. Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-I and IGF-binding protein-3. *J. Natl Cancer Inst.* 91:620–625.
24. Chan, J. M., M. J. Stampfer, E. Giovannucci, P. H. Gann, J. Ma, P. Wilkinson, et al. 1998. Plasma insulin-like growth factor-I and prostate cancer risk: a prospective study. *Science* 279:563–566.
25. Brooks, A. J., and M. J. Waters. 2010. The growth hormone receptor: mechanism of activation and clinical implications. *Nat. Rev. Endocrinol.* 6:515–525.
26. Laron, Z. 2015. Lesson from 50 years of study of laron syndrome. *Endocr. Pra.* 12:1395–1402.
27. NIH National Cancer Institute. Available from: <http://dceg.cancer.gov/research/what-we-study/second-cancers> (accessed date: 22 February 2015).
28. Sachdev, D., and D. Yee. 2001. The IGF system and breast cancer. *Endocr. Relat. Cancer* 8:197–209.
29. Sara, V. R., and K. Hall. 1990. Insulin-like growth factors and their binding proteins. *Physiol. Rev.* 70:591–614.
30. Rosenfeld, R. G., G. Lamson, H. Pham, Y. Oh, C. Conover, D. D. De Leon, et al. 1990. Insulinlike growth factor-binding proteins. *Recent Prog. Horm. Res.* 46:99–159; discussion -63.
31. El-Shewy, H. M., M. H. Lee, L. M. Obeid, A. A. Jaffa, and L. M. Luttrell. 2007. The insulin-like growth factor type 1 and insulin-like growth factor type 2/mannose-6-phosphate receptors independently regulate ERK1/2 activity in HEK293 cells. *J. Biol. Chem.* 282:26150–26157.
32. Bach, L. A., and L. J. Hale. 2015. Insulin-like growth factors and kidney disease. *Am. J. Kidney Dis.* 65:327–336.
33. Kaplan, R. C., H. D. Strickler, T. E. Rohan, R. Muzumdar, and D. L. Brown. 2005. Insulin-like growth factors and coronary heart disease. *Cardiol. Rev.* 13:35–39.
34. Stewart, C. E., and P. Rotwein. 1996. Growth, differentiation, and survival: multiple physiological functions for insulin-like growth factors. *Physiol. Rev.* 76:1005–1026.
35. Yu, H., and T. Rohan. 2000. Role of the insulin-like growth factor family in cancer development and progression. *J. Natl Cancer Inst.* 92:1472–1489.
36. Pollak, M. N., E. S. Schernhammer, and S. E. Hankinson. 2004. Insulin-like growth factors and neoplasia. *Nat. Rev. Cancer* 4:505–518.
37. Hartog, H., H. M. Boezen, M. M. de Jong, M. Schaapveld, J. Wesseling, and W. T. van der Graaf. 2013. Prognostic value of insulin-like growth factor 1 and insulin-like growth factor binding protein 3 blood levels in breast cancer. *Breast* 22:1155–1160.
38. Yakar, S., D. Leroith, and P. Brodt. 2005. The role of the growth hormone/insulin-like growth factor axis in tumor growth and progression: lessons from animal models. *Cytokine Growth Factor Rev.* 16(4–5):407–420.
39. Laron, Z. 2001. Insulin-like growth factor 1 (IGF-1): a growth hormone. *Mol. Pathol.* 54:311–316.
40. Sklar, C. A., A. C. Mertens, P. Mitby, G. Occhiogrosso, J. Qin, G. Heller, et al. 2002. Risk of disease recurrence and second neoplasms in survivors of childhood cancer treated with growth hormone: a report from the childhood cancer survivor study. *J. Clin. Endocrinol. Metab.* 87:3136–3141.
41. Peyrat, J. P., J. Bonnetterre, B. Hecquet, P. Vennin, M. M. Louchez, C. Fournier, et al. 1993. Plasma insulin-like growth factor-I (IGF-I) concentrations in human breast cancer. *Eur. J. Cancer* 29A:492–497.
42. Endogenous, H., Breast Cancer Collaborative G; T. J. Key, P. N. Appleby, G. K. Reeves, and A. W. Roddam. 2010. Insulin-like growth factor 1 (IGF1), IGF binding protein 3 (IGFBP3), and breast cancer risk: pooled individual data analysis of 17 prospective studies. *Lancet Oncol.* 11:530–542.
43. Rinaldi, S., P. H. Peeters, F. Berrino, L. Dossus, C. Biessy, A. Olsen, et al. 2006. IGF-I, IGFBP-3 and breast cancer risk in women: the European Prospective Investigation into Cancer and Nutrition (EPIC). *Endocr. Relat. Cancer* 13:593–605.
44. Kaaks, R., T. Johnson, K. Tikk, D. Sookthai, A. Tjonneland, N. Roswall, et al. 2014 Jun. Insulin-like growth factor I and risk of breast cancer by age and hormone receptor status-A prospective study within the EPIC cohort. *Int. J. Cancer* 134:2683–2690.
45. Baglietto, L., D. R. English, J. L. Hopper, H. A. Morris, W. D. Tilley, and G. G. Giles. 2007. Circulating insulin-like growth factor-I and binding protein-3 and the risk of breast cancer. *Cancer Epidemiol. Biomark. Prev.* 16:763–768.
46. Renehan, A. G., M. Zwahlen, C. Minder, S. T. O'Dwyer, S. M. Shalet, and M. Egger. 2004. Insulin-like growth factor (IGF)-I, IGF binding protein-3, and

- cancer risk: systematic review and meta-regression analysis. *Lancet* 363:1346–1353.
47. Shi, R., H. Yu, J. McLarty, and J. Glass. 2004. IGF-I and breast cancer: a meta-analysis. *Int. J. Cancer* 111:418–423.
 48. Sugumar, A., Y. C. Liu, Q. Xia, Y. S. Koh, and K. Matsuo. 2004. Insulin-like growth factor (IGF)-I and IGF-binding protein 3 and the risk of premenopausal breast cancer: a meta-analysis of literature. *Int. J. Cancer* 111:293–297.
 49. Schernhammer, E. S., J. M. Holly, D. J. Hunter, M. N. Pollak, and S. E. Hankinson. 2006. Insulin-like growth factor-I, its binding proteins (IGFBP-1 and IGFBP-3), and growth hormone and breast cancer risk in the nurses health study II. *Endocr. Relat. Cancer* 13:583–592.
 50. Kaaks, R., E. Lundin, S. Rinaldi, J. Manjer, C. Biessy, S. Soderberg, et al. 2002. Prospective study of IGF-I, IGF-binding proteins, and breast cancer risk, in northern and southern Sweden. *Cancer Causes Control* 13:307–316.
 51. Maor, S., D. Mayer, R. I. Yarden, A. V. Lee, R. Sarfstein, H. Werner, et al. 2006. Estrogen receptor regulates insulin-like growth factor-I receptor gene expression in breast tumor cells: involvement of transcription factor Sp1. *J. Endocrinol.* 191:605–612.
 52. Stewart, A. J., M. D. Johnson, F. E. May, and B. R. Westley. 1990. Role of insulin-like growth factors and the type I insulin-like growth factor receptor in the estrogen-stimulated proliferation of human breast cancer cells. *J. Biol. Chem.* 265:21172–21178.
 53. Ahn, J., S. J. Weinstein, K. Snyder, M. N. Pollak, J. Virtamo, and D. Albanes. 2006. No association between serum insulin-like growth factor (IGF)-I, IGF-binding protein-3, and lung cancer risk. *Cancer Epidemiol. Biomark. Prev.* 15:2010–2012.
 54. Spitz, M. R., M. J. Barnett, G. E. Goodman, M. D. Thornquist, X. Wu, and M. Pollak. 2002. Serum insulin-like growth factor (IGF) and IGF-binding protein levels and risk of lung cancer: a case-control study nested in the beta-carotene and retinol efficacy trial cohort. *Cancer Epidemiol. Biomark. Prev.* 11:1413–1418.
 55. London, S. J., J. M. Yuan, G. S. Travlos, Y. T. Gao, R. E. Wilson, R. K. Ross, et al. 2002. Insulin-like growth factor I, IGF-binding protein 3, and lung cancer risk in a prospective study of men in China. *J. Natl Cancer Inst.* 94:749–754.
 56. Lukanova, A., P. Toniolo, A. Akhmedkhanov, C. Biessy, N. J. Haley, R. E. Shore, et al. 2001. A prospective study of insulin-like growth factor-I, IGF-binding proteins-1, -2 and -3 and lung cancer risk in women. *Int. J. Cancer* 92:888–892.
 57. Morris, J. K., L. M. George, T. Wu, and N. J. Wald. 2006. Insulin-like growth factors and cancer: no role in screening. Evidence from the BUPA study and meta-analysis of prospective epidemiological studies. *Br. J. Cancer* 95:112–117.
 58. Chen, B., S. Liu, W. Xu, X. Wang, W. Zhao, and J. Wu. 2009. IGF-I and IGFBP-3 and the risk of lung cancer: a meta-analysis based on nested case-control studies. *J. Exp. Clin. Cancer Res.* 28:89.
 59. Cao, H., G. Wang, L. Meng, H. Shen, Z. Feng, Q. Liu, et al. 2012. Association between circulating levels of IGF-1 and IGFBP-3 and lung cancer risk: a meta-analysis. *PLoS ONE* 7:e49884.
 60. Jaques, G., K. Noll, B. Wegmann, S. Witten, E. Kogan, R. T. Radulescu, et al. 1997. Nuclear localization of insulin-like growth factor binding protein 3 in a lung cancer cell line. *Endocrinology* 138:1767–1770.
 61. Favoni, R. E., A. de Cupis, F. Ravera, C. Cantoni, P. Pirani, A. Ardizzoni, et al. 1994. Expression and function of the insulin-like growth factor I system in human non-small-cell lung cancer and normal lung cell lines. *Int. J. Cancer* 56:858–866.
 62. Cohen, P., D. M. Peehl, T. A. Stamey, K. F. Wilson, D. R. Clemmons, and R. G. Rosenfeld. 1993. Elevated levels of insulin-like growth factor-binding protein-2 in the serum of prostate cancer patients. *J. Clin. Endocrinol. Metab.* 76:1031–1035.
 63. Kanety, H., Y. Madjar, Y. Dagan, J. Levi, M. Z. Papa, C. Pariente, et al. 1993. Serum insulin-like growth factor-binding protein-2 (IGFBP-2) is increased and IGFBP-3 is decreased in patients with prostate cancer: correlation with serum prostate-specific antigen. *J. Clin. Endocrinol. Metab.* 77:229–233.
 64. Mantzoros, C. S., A. Tzonou, L. B. Signorello, M. Stampfer, D. Trichopoulos, and H. O. Adami. 1997. Insulin-like growth factor 1 in relation to prostate cancer and benign prostatic hyperplasia. *Br. J. Cancer* 76:1115–1118.
 65. Dunn, S. E., F. W. Kari, J. French, J. R. Leininger, G. Travlos, R. Wilson, et al. 1997. Dietary restriction reduces insulin-like growth factor I levels, which modulates apoptosis, cell proliferation, and tumor progression in p53-deficient mice. *Cancer Res.* 57:4667–4672.
 66. Pollak, M., W. Beamer, and J. C. Zhang. 1998. Insulin-like growth factors and prostate cancer. *Cancer Metastasis Rev.* 17:383–390.
 67. Saikali, Z., H. Setya, G. Singh, and S. Persad. 2008. Role of IGF-1/IGF-1R in regulation of invasion in DU145 prostate cancer cells. *Cancer Cell Int.* 8:10.
 68. Grimberg, A., and P. Cohen. 1999. Growth hormone and prostate cancer: guilty by association? *J. Endocrinol. Invest.* 22(5 Suppl):64–73.
 69. Nomura, A. M., G. N. Stemmermann, J. Lee, and M. N. Pollak. 2003. Serum insulin-like growth factor I and

- subsequent risk of colorectal cancer among Japanese-American men. *Am. J. Epidemiol.* 158:424–431.
70. Palmqvist, R., G. Hallmans, S. Rinaldi, C. Biessy, R. Stenling, E. Riboli, et al. 2002. Plasma insulin-like growth factor 1, insulin-like growth factor binding protein 3, and risk of colorectal cancer: a prospective study in northern Sweden. *Gut* 50:642–646.
 71. Tripkovic, I., A. Tripkovic, M. Strnad, V. Capkun, and L. Zekan. 2007. Role of insulin-like growth factor-1 in colon cancerogenesis: a case-control study. *Arch. Med. Res.* 38:519–525.
 72. Kaaks, R., P. Toniolo, A. Akhmedkhanov, A. Lukanova, C. Biessy, H. Dechaud, et al. 2000. Serum C-peptide, insulin-like growth factor (IGF)-I, IGF-binding proteins, and colorectal cancer risk in women. *J. Natl Cancer Inst.* 92:1592–1600.
 73. Giovannucci, E., M. N. Pollak, E. A. Platz, W. C. Willett, M. J. Stampfer, N. Majeed, et al. 2000. A prospective study of plasma insulin-like growth factor-1 and binding protein-3 and risk of colorectal neoplasia in women. *Cancer Epidemiol. Biomark. Prev.* 9:345–349.
 74. Suzuki, S., M. Kojima, S. Tokudome, K. Suzuki, K. Ozasa, Y. Ito, et al. 2009. Insulin-like growth factor (IGF)-I, IGF-II, IGF binding protein-3, and risk of colorectal cancer: a nested case-control study in the Japan Collaborative Cohort study. *Asian Pac. J. Cancer Prev.* 10(Suppl):45–49.
 75. Max, J. B., P. J. Limburg, A. Ogunseitan, R. Z. Stolzenberg-Solomon, R. A. Vierkant, M. J. Pollak, et al. 2008. IGF-I, IGFBP-3, and IGF-I/IGFBP-3 ratio: no association with incident colorectal cancer in the alpha-tocopherol, beta-carotene cancer prevention study. *Cancer Epidemiol. Biomark. Prev.* 17:1832–1834.
 76. Peters, G., S. Gongoll, C. Langner, M. Mengel, P. Piso, J. Klempnauer, et al. 2003. IGF-1R, IGF-1 and IGF-2 expression as potential prognostic and predictive markers in colorectal-cancer. *Virchows Arch.* 443:139–145.
 77. Freier, S., O. Weiss, M. Eran, A. Flyvbjerg, R. Dahan, I. Nephesh, et al. 1999. Expression of the insulin-like growth factors and their receptors in adenocarcinoma of the colon. *Gut* 44:704–708.
 78. Hakam, A., T. J. Yeatman, L. Lu, L. Mora, G. Marcet, S. V. Nicosia, et al. 1999. Expression of insulin-like growth factor-1 receptor in human colorectal cancer. *Hum. Pathol.* 30:1128–1133.
 79. Shiratsuchi, I., Y. Akagi, A. Kawahara, T. Kinugasa, K. Romeo, T. Yoshida, et al. 2011. Expression of IGF-1 and IGF-1R and their relation to clinicopathological factors in colorectal cancer. *Anticancer Res.* 31:2541–2545.
 80. Orme, S. M., R. J. McNally, R. A. Cartwright, and P. E. Belchetz. 1998. Mortality and cancer incidence in acromegaly: a retrospective cohort study. United Kingdom Acromegaly study group. *J. Clin. Endocrinol. Metab.* 83:2730–2734.
 81. Jenkins, P. J., V. Frajese, A. M. Jones, C. Camacho-Hubner, D. G. Lowe, P. D. Fairclough, et al. 2000. Insulin-like growth factor I and the development of colorectal neoplasia in acromegaly. *J. Clin. Endocrinol. Metab.* 85:3218–3221.
 82. Zecha, H., H. P. Schmid, A. Tschopp, T. Sulser, and D. S. Engeler. 2011. High incidence of independent second malignancies after non-muscle-invasive bladder cancer. *Scand. J. Urol. Nephrol.* 45:245–250.
 83. Ge, J., H. F. Gou, Y. Chen, K. Cheng, L. H. Li, H. Dong, et al. 2013. Clinical characteristics of patients with solitary pulmonary mass after radical treatment for primary cancers: pulmonary metastasis or second primary lung cancer? *Cancer Invest.* 31:397–403.
 84. Brenner, D. J. 2010. Contralateral second breast cancers: prediction and prevention. *J. Natl Cancer Inst.* 102:444–445.
 85. Rusner, C., K. Wolf, U. Bandemer-Greulich, J. Engel, C. Stegmaier, B. Holleczer, et al. 2014. Risk of contralateral second primary breast cancer according to hormone receptor status in Germany. *Breast Cancer Res.* 16:452.
 86. Crump, M., and D. Hodgson. 2009. Secondary breast cancer in Hodgkin's lymphoma survivors. *J. Clin. Oncol.* 27:4229–4231.
 87. Bhatia, S., L. L. Robison, O. Oberlin, M. Greenberg, G. Bunin, F. Fossati-Bellani, et al. 1996. Breast cancer and other second neoplasms after childhood Hodgkin's disease. *N. Engl. J. Med.* 334:745–751.
 88. Swerdlow, A. J., R. Cooke, A. Bates, D. Cunningham, S. J. Falk, D. Gilson, et al. 2012. Breast cancer risk after supradiaphragmatic radiotherapy for Hodgkin's lymphoma in England and Wales: a National Cohort Study. *J. Clin. Oncol.* 30:2745–2752.
 89. Cooke, R., M. E. Jones, D. Cunningham, S. J. Falk, D. Gilson, B. W. Hancock, et al. 2013. Breast cancer risk following Hodgkin lymphoma radiotherapy in relation to menstrual and reproductive factors. *Br. J. Cancer* 108:2399–2406.
 90. Bernstein, J. L., R. H. Lapinski, S. S. Thakore, J. T. Doucette, and W. D. Thompson. 2003. The descriptive epidemiology of second primary breast cancer. *Epidemiology* 14:552–558.
 91. Storm, H. H., and O. M. Jensen. 1986. Risk of contralateral breast cancer in Denmark 1943–80. *Br. J. Cancer* 54:483–492.
 92. Hancock, S. L., M. A. Tucker, and R. T. Hoppe. 1993. Breast cancer after treatment of Hodgkin's disease. *J. Natl Cancer Inst.* 85:25–31.
 93. Kleinberg, D. L., M. Feldman, and W. Ruan. 2000. IGF-I: an essential factor in terminal end bud formation and ductal morphogenesis. *J. Mammary Gland Biol. Neoplasia* 5:7–17.

94. Frasca, F., G. Pandini, L. Sciacca, V. Pezzino, S. Squatrito, A. Belfiore, et al. 2008. The role of insulin receptors and IGF-I receptors in cancer and other diseases. *Arch. Physiol. Biochem.* 114:23–37.
95. Yee, D., S. Paik, G. S. Lebovic, R. R. Marcus, R. E. Favoni, K. J. Cullen, et al. 1989. Analysis of insulin-like growth factor I gene expression in malignancy: evidence for a paracrine role in human breast cancer. *Mol. Endocrinol.* 3:509–517.
96. Allen, N. E., A. W. Roddam, D. S. Allen, I. S. Fentiman, Silva. I. Dos Santos, J. Peto, et al. 2005. A prospective study of serum insulin-like growth factor-I (IGF-I), IGF-II, IGF-binding protein-3 and breast cancer risk. *Br. J. Cancer* 92:1283–1287.
97. Smith, J., D. Axelrod, B. Singh, and D. Kleinberg. 2011. Prevention of breast cancer: the case for studying inhibition of IGF-1 actions. *Ann. Oncol.* 22(Suppl 1):i50–i52.
98. Coyte, A., D. S. Morrison, and P. McLoone. 2014. Second primary cancer risk - the impact of applying different definitions of multiple primaries: results from a retrospective population-based cancer registry study. *BMC Cancer* 14:272.
99. Kaufman, E. L., J. S. Jacobson, D. L. Hershman, M. Desai, and A. I. Neugut. 2008. Effect of breast cancer radiotherapy and cigarette smoking on risk of second primary lung cancer. *J. Clin. Oncol.* 26:392–398.
100. Senthil, S., C. J. Haasbeek, F. J. Lagerwaard, W. F. Verbakel, P. F. de Haan, B. J. Slotman, et al. 2013. Radiotherapy for a second primary lung cancer arising post-pneumonectomy: planning considerations and clinical outcomes. *J. Thorac. Dis.* 5:116–122.
101. Argiris, A., B. E. Brockstein, D. J. Haraf, K. M. Stenson, B. B. Mittal, M. S. Kies, et al. 2004. Competing causes of death and second primary tumors in patients with locoregionally advanced head and neck cancer treated with chemoradiotherapy. *Clin. Cancer Res.* 10:1956–1962.
102. Boyle, J. M., D. J. Tandberg, J. P. Chino, T. A. D'Amico, N. E. Ready, and C. R. Kelsey. 2015. Smoking history predicts for increased risk of second primary lung cancer: a comprehensive analysis. *Cancer* 121:598–604.
103. Richardson, G. E., M. A. Tucker, D. J. Venzon, R. I. Linnoila, R. Phelps, J. C. Phares, et al. 1993. Smoking cessation after successful treatment of small-cell lung cancer is associated with fewer smoking-related second primary cancers. *Ann. Intern. Med.* 119:383–390.
104. Johnson, B. E., P. Cortazar, and J. P. Chute. 1997. Second lung cancers in patients successfully treated for lung cancer. *Semin. Oncol.* 24:492–499.
105. Schonfeld, S. J., R. E. Curtis, W. F. Anderson, and A. Berrington de Gonzalez. 2012. The risk of a second primary lung cancer after a first invasive breast cancer according to estrogen receptor status. *Cancer Causes Control* 23:1721–1728.
106. Kim, W. Y., M. J. Kim, H. Moon, P. Yuan, J. S. Kim, J. K. Woo, et al. 2011. Differential impacts of insulin-like growth factor-binding protein-3 (IGFBP-3) in epithelial IGF-induced lung cancer development. *Endocrinology* 152:2164–2173.
107. Levi, F., L. Randimbison, V. C. Te, G. Erler, and C. La Vecchia. 1999. Second primary tumors after prostate carcinoma. *Cancer* 86:1567–1570.
108. Davis, E. J., J. L. Beebe-Dimmer, C. L. Yee, and K. A. Cooney. 2014. Risk of second primary tumors in men diagnosed with prostate cancer: a population-based cohort study. *Cancer* 120:2735–2741.
109. Abern, M. R., A. M. Dude, M. Tsivian, and C. L. Coogan. 2013. The characteristics of bladder cancer after radiotherapy for prostate cancer. *Urologic Oncol.* 31:1628–1634.
110. Anderson, J. K., S. Alanee, B. Lindgren, and J. Slaton. 2013. The risk of bladder cancer in patients diagnosed with other primary neoplasms: analysis of the SEER database. *Urologic Oncol.* 31:862–865.
111. Braisch, U., M. Meyer, and M. Radespiel-Troger. 2012. Risk of subsequent primary cancer among prostate cancer patients in Bavaria, Germany. *Eur. J. Cancer Prev.* 21:552–559.
112. Zhang, H., J. L. Bermejo, J. Sundquist, and K. Hemminki. 2009. Prostate cancer as a first and second cancer: effect of family history. *Br. J. Cancer* 101:935–939.
113. Chrouser, K., B. Leibovich, E. Bergstralh, H. Zincke, and M. Blute. 2005. Bladder cancer risk following primary and adjuvant external beam radiation for prostate cancer. *J. Urol.* 174:107–110; discussion 10–1.
114. Kok, D. E., S. A. van de Schans, L. Liu, E. Kampman, J. W. Coebergh, L. A. Kiemeny, et al. 2013. Risk of prostate cancer among cancer survivors in the Netherlands. *Cancer Epidemiol.* 37:140–145.
115. Kellen, E., M. P. Zeegers, M. Dirx, S. Housterman, J. Droste, G. Lawrence, et al. 2007. Occurrence of both bladder and prostate cancer in five cancer registries in Belgium, The Netherlands and the United Kingdom. *Eur. J. Cancer* 43:1694–1700.
116. Kurokawa, K., K. Ito, T. Yamamoto, H. Takechi, S. Miyamoto, K. Suzuki, et al. 2004. Comparative study on the prevalence of clinically detectable prostate cancer in patients with and without bladder cancer. *Urology* 63:268–272.
117. Bradford, P. T., D. M. Freedman, A. M. Goldstein, and M. A. Tucker. 2010. Increased risk of second primary cancers after a diagnosis of melanoma. *Arch. Dermatol.* 146:265–272.
118. Chen, C., Q. Zhang, S. Liu, K. R. Parajuli, Y. Qu, J. Mei, et al. 2015. IL-17 and insulin/IGF1 enhance

- adhesion of prostate cancer cells to vascular endothelial cells through CD44-VCAM-1 interaction. *Prostate* 75:883–895.
119. Pickles, T., and N. Phillips. 2002. The risk of second malignancy in men with prostate cancer treated with or without radiation in British Columbia, 1984–2000. *Radiother. Oncol.* 65:145–151.
 120. McCredie, M., G. J. Macfarlane, J. Stewart, and M. Coates. 1996. Second primary cancers following cancers of the kidney and prostate in New South Wales (Australia), 1972–91. *Cancer Causes Control* 7:337–344.
 121. Brenner, D. J., R. E. Curtis, E. J. Hall, and E. Ron. 2000. Second malignancies in prostate carcinoma patients after radiotherapy compared with surgery. *Cancer* 88:398–406.
 122. Baxter, N. N., J. E. Tepper, S. B. Durham, D. A. Rothenberger, and B. A. Virnig. 2005. Increased risk of rectal cancer after prostate radiation: a population-based study. *Gastroenterology* 128:819–824.
 123. Chaturvedi, A. K. E., E. A. Engels, E. S. Gilbert, B. E. Chen, H. Storm, C. F. Lynch, et al. 2007. Second cancers among 104 760 survivors of cervical cancer: evaluation of long-term risk. *J. Natl Cancer Inst.* 99:1634–1643.
 124. Vigneri, P. G., E. Tirro, M. S. Pennisi, M. Massimino, S. Stella, C. Romano, et al. 2015. The Insulin/IGF System in colorectal cancer development and resistance to therapy. *Front. Oncol.* 5:230.
 125. Grannis, F. W. Jr. 2013. Minimizing over-diagnosis in lung cancer screening. *J. Surg. Oncol.* 108:289–293.
 126. Tworoger, S. S., and S. E. Hankinson. 2006. Use of biomarkers in epidemiologic studies: minimizing the influence of measurement error in the study design and analysis. *Cancer Causes Control* 17:889–899.
 127. LeRoith, D. 2012. Insulin-like growth factors and cancer: from basic biology to therapeutics. *Development CDDa*, editor: Springer, Springer US.
 128. Bokarewa, M. I., M. C. Erlandsson, J. Bjersing, M. Dehlin, and K. Mannerkorpi. 2014. Smoking is associated with reduced leptin and neuropeptide Y levels and higher pain experience in patients with fibromyalgia. *Mediators Inflamm.* 2014:627041.
 129. Lin, I. H., M. L. Ho, H. Y. Chen, H. S. Lee, C. C. Huang, Y. H. Chu, et al. 2012. Smoking, green tea consumption, genetic polymorphisms in the insulin-like growth factors and lung cancer risk. *PLoS ONE* 7:e30951.
 130. Schisterman, E. F., S. R. Cole, and R. W. Platt. 2009. Overadjustment bias and unnecessary adjustment in epidemiologic studies. *Epidemiology* 20:488–495.